



## Integrated Microchip Sensors for Detecting *Escherichia coli* O157:H7 and *Salmonella enterica* Based on TaqMan Assays


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Oct. 2009

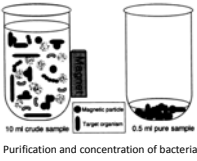



## Microfluidic Sensor based on Manipulation of Magnetic Beads

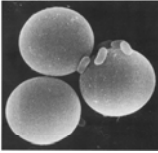
- Integration of multiple steps for PCR based detection
  - Bacterial cell capture by immunomagnetic separation (off-chip)
  - Cell-bead complex concentration by magnetic field
  - Electric lysis of cells and release of plasmid DNA
  - Adsorption of plasmid DNA on packed bed of magnetic beads
  - TaqMan PCR assay (off-chip)



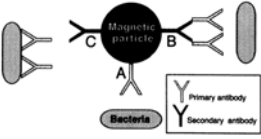
## Immunomagnetic Separation (IMS) for Bacteria Separation from Food Matrix



Purification and concentration of bacteria




Scanning electron microscopy image of bacteria bound to Dynabeads M450 coated with antibody

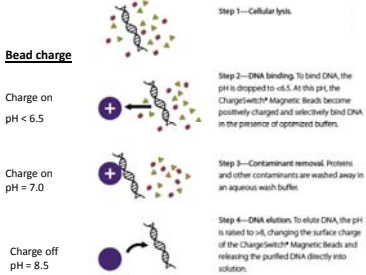


Dynabeads® anti-Salmonella

Popovic T., Skjerve E., Cudjoe K.S., Homes E., Ugelstad J., Uhlen M. *Clinical Microbiology Review* 7 (1994) 43-54.




## ChargeSwitch® Magnetic Beads for Plasmid DNA Collection

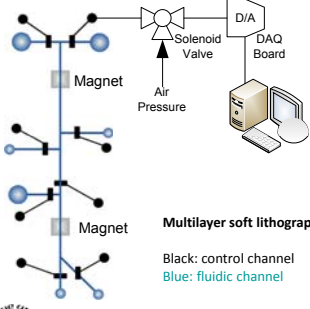


Bead charge

- Charge on pH < 6.5
- Charge on pH = 7.0
- Charge off pH = 8.5





## Microchip Design



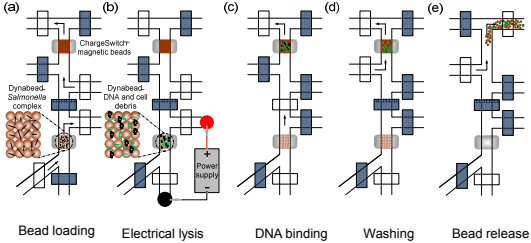
Fully closed valve (Valve closing for rounded channel)

Multilayer soft lithography


Black: control channel  
Blue: fluidic channel

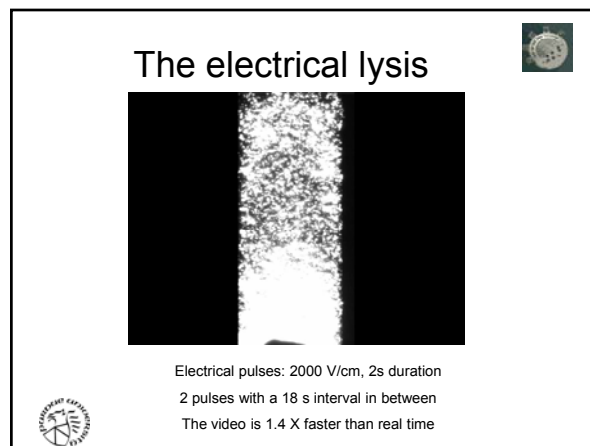
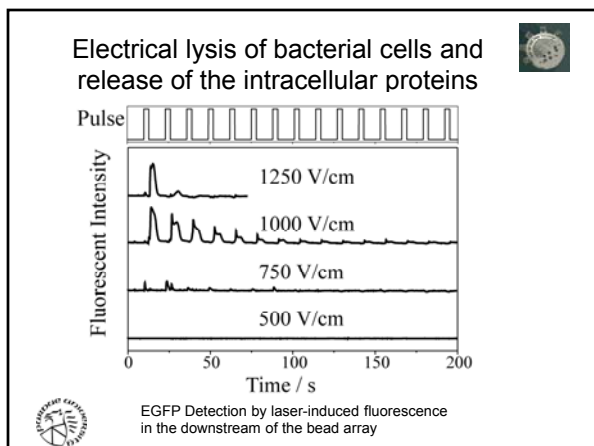
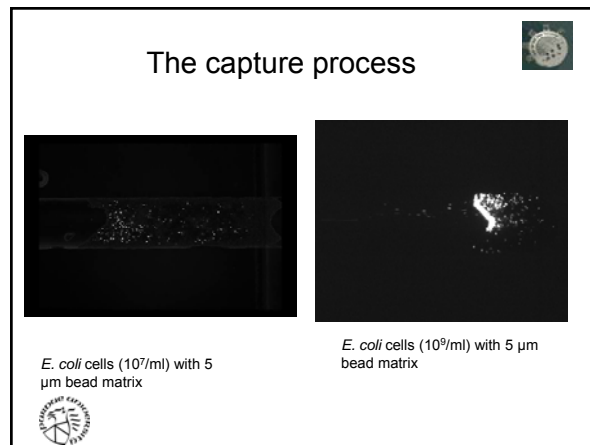
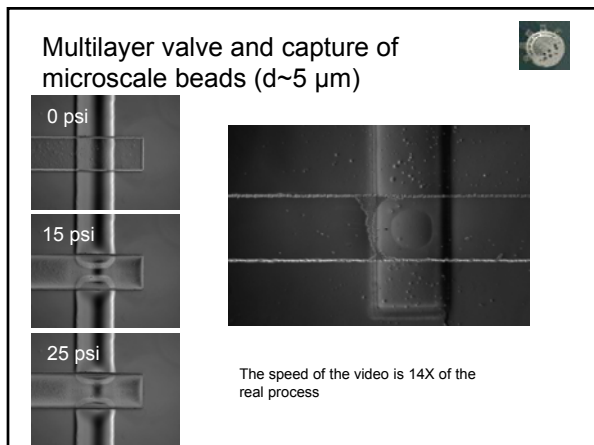
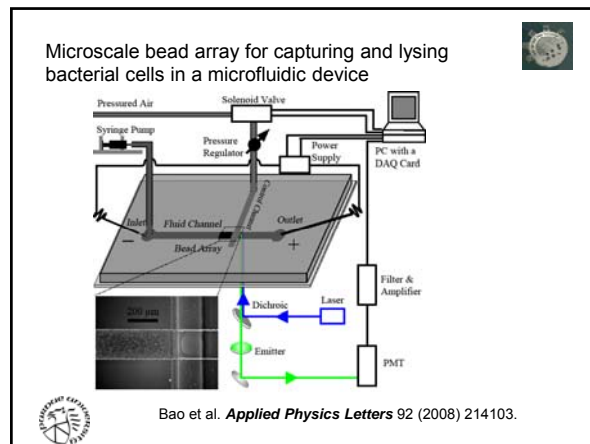
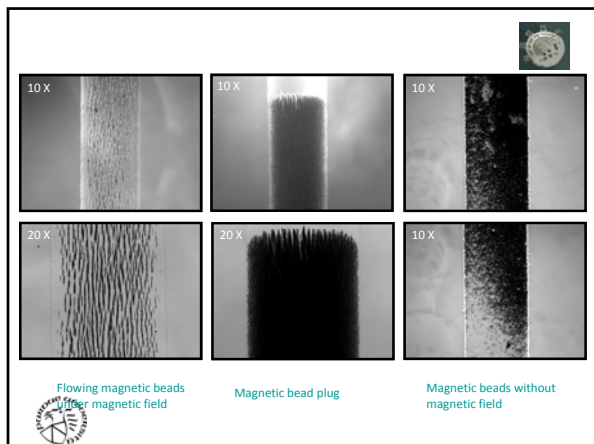



## On-chip Cell Concentration, Lysis and DNA Purification

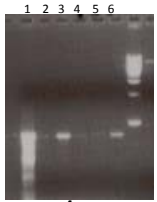


(a) Bead loading    (b) Electrical lysis    (c) DNA binding    (d) Washing    (e) Bead release





### Preliminary Results



1 2 3 4 5 6

**envA gene**

**PCR conditions:**  
initially at 94 °C for 1 min, 29 cycles at 94 °C for 1 min, at 50 °C for 1 min, at 72 °C for 1 min, and a final extension at 72 °C for 10 min

**Primer:**  
Forward:  
CGGTGGTTTTAAGCGTACTCTT  
Reverse :  
CGAATATGCTCCACAAGTTA

**Lane 1:** Positive control, cells were lysed by heating for 10 min at 90 °C, and DNA were not purified. ~10<sup>5</sup> cells


**Lane 2:** 1500 V/cm (10 s interval and 1 s pulse for 3 min), ~10<sup>6</sup> cells

**Lane 3:** 1200 V/cm (10 s interval and 1 s pulse for 3 min), ~10<sup>6</sup> cells

**Lane 4:** 1000 V/cm (10 s interval and 1 s pulse for 3 min), ~10<sup>4</sup> cells

**Lane 5:** 1000 V/cm (10 s interval and 1 s pulse for 3 min), ~10<sup>4</sup> cells, from washing of the chargeswitch beads

**Lane 6:** 1000 V/cm (10 s interval and 1 s pulse for 3 min), ~10<sup>7</sup> cells





### Current Challenges and Future Work

**Problems:**

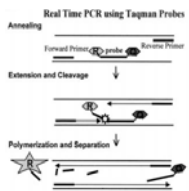
1. Unknown cell number in each experimental run.
2. Low sensitivity.
3. DNA desorption from chargeswitch beads.

**Future work:**

1. Apply cells expressing fluorescent protein (e.g. GFP) for enumeration of cells.
2. Pinpoint the bottleneck for improving sensitivity among several effects: completeness of electric lysis release, DNA capture efficiency, DNA adsorption on IMS beads, PCR condition
3. Apply RT-PCR.
4. Reduce the flow rate of wash buffer, and increase the density of ChargeSwitch<sup>®</sup> magnetic beads.




### Real-Time PCR Assay



**Real Time PCR using Taqman Probes**



1. Probes and primers anneal to target sequence. Taqman probes have two covalently linked fluorescent dyes: a reporter (R) and a quencher (Q). On the probe, the reporter dye emission is quenched.
2. During each extension cycle, the 5'→3' exonuclease activity of Taq DNA polymerase cleaves the reporter dye from the probe.
3. Once separated from the quencher, the reporter dye emits its characteristic fluorescence, which is measured in every cycle by a detector.

TaqMan<sup>®</sup> Pathogen Detection Kits (Applied Biosystems)  
Primers and Taqman Probes: Applied Biosystems Primer Express s

### Conclusions

- Magnetic beads are manipulated by a magnet in a microfluidic channel to form a closely packed bed
- Plasmid DNA can be released from *Salmonella* cells by electric lysis
- PCR is conducted based on extracted DNA from bacteria

### Acknowledgements

- Dr. Ning Bao, Tao Geng, Dr. Kirshna Mishra
- Funding from ARS-CFSE

